

Original article

Effect of temperature on microbial changes during kimchi fermentation

F. Tabatabaei Yazdi*, B. Alizadeh Behbahani, M. Mohebbi, A. Mortazavi, A. Ghaitaranpour Department of Food Science and Technology, Faculty of Agriculture, Ferdowsi University of Mashhad, Iran.

*Corresponding author; Department of Food Science and Technology, Faculty of Agriculture, Ferdowsi University of Mashhad, Iran.

ARTICLE INFO

Article history:
Received 10 January 2013
Accepted 22 January 2013
Available online 23 January 2013

Keywords:
Kimchi
Micro flora
Temperature
Yeast

ABSTRACT

Kimchi, a salt-pickled and fermented vegetable, is one of the most popular foods in Asian countries, such as Korea, and more and more people in the world get used to its taste. In this study, Kimchi were fermented at two different temperatures. After acidification to pH=4.2, LAB were isolated, enumerated. The purpose of this study is to determine the amount of micro flora of Kimchi. The results show that the naturally occurring lactic acid bacteria load was found to vary between 1.5×10⁴ cfu/gr to 1.9×10⁸ cfu/gr at 20° c. The yeast counts increased from 1.13×10⁴ cfu/gr to 5×10⁷ cfu/gr at 20 c°. The LAB counts on MRS increased from 1.5×10⁴ cfu/gr to 6.1×10⁸ cfu/gr at 30 °c. The largest increase in the numbers of LAB was noted during the first 24 h of fermentation and further incubation led to decrease. The yeast counts increased from 1.13×10^4 cfu/gr to 5.1×107 cfu/gr. Maximum total acids produced in kimchi at 30°C. At 30°C, the optimum-ripening period was 1 day and the edible period was also 1-2 days. The results show that the temperature is effective on microbial changes during kimchi fermentation.

© 2013 Sjournals. All rights reserved.

1. Introduction

The word kimchi is the generic term for Korean fermented vegetables, which is derived from the Chinese characters pronounced chimchae, meaning brined vegetables Kimchi is a Korean traditional fermented vegetable. Most kimchi is characterized by its hot taste due to the use of fairly high amount of chili pepper and its visibility in

the product. However, some kimchi are made without chili pepper, but with garlic and ginger together with vegetables and other ingredients. Traditionally, kimchi was made in every household in Korean rural areas to provide the vegetables for the winter when other fresh vegetables are not readily available (hand). Kimchi contains various health-promoting components, including β-carotene, chlorophyll, vitamin C, and dietary fiber (Park et al., 1995). It has its best flavor, taste, and texture when optimally fermented at about pH 4.2; after optimum fermentation, Kimchi quality deteriorates rapidly due to formation of excessive organic acids and texture softening (Kim and Kim, 2003). Many types of kimchi are available depending on the raw ingredients used, processing methods, harvest seasons, and geographical regions. Although baechu (Korean cabbage) and radish are the most widely used in the making of kimchi, other vegetables are used depending on their seasonal availability. Because of the cold northern, and the mild southern Korean winters, the kimchi prepared in the north contains less salt, whereas the kimchi in the south requires more salt for long-term preservation. Also, people living near the sea naturally use more fish products in their kimchi(Lee et al., 1997). Therefore, the control of the fermentation process is needed to preserve quality of Kimchi and to extend its shelf-life. Kimchi fermentation occurs mainly by the microorganisms naturally present in the raw materials that contain numerous microflora including LAB. Various LABs may initiate fermentation, but hetero fermentative type LAB increased rapidly with organic acids accumulation and homo fermentative type LAB increased thereafter. Many homo and hetero fermentative LAB, such as genus Lactobacillus, Leuconostoc, Weissella, Lactococcus, Streptococcus and Pediococcus were isolated and identified from various kimchi samples, but genus Leuconostoc, Weissella and other LAB producing organic acids, carbon dioxide and bacteriocins are the most important microorganisms for the controlled fermentation of kimchi (Lee etal., 2002). Numerous chemical, physical, and biological factors may contribute directly to the growth of microorganisms and the extent of fermentation. Kimchi has a low caloric content, but it is rich in minerals, vitamins, and dietary fiber. The protein and lipid contents can be increased with various subingredients like fish, clams, oysters, and meat, which give kimchi its characteristic savor. Traditionally in Korea, large quantities of kimchi are prepared as an annual event, kimjang, for eating during the winter when the fresh vegetable supply is limited. Most of the vegetables cultivated in Korea are used as sources for making kimchi. Although 161 or 187 kinds of kimchi are currently reported (Edward, 2008). The important factors that affect kimchi fermentation are microorganisms, temperature, salt concentration, fermentable carbohydrates, other available nutrients, or any inhibitory compounds in raw materials used, as well as oxygen and pH.

The purpose of this study was to determine the amount of micro flora of kimchi. Another purpose is studying Changes in microbiological composition of kimchi during fermentation.

2. Materials and methods

In this study, the ingredients (Chinese cabbage, Leek, red pepper powder, Garlic, Ginger, Sugar, rice flour) used in kimchi preparation were purchased from local markets in Mashhad, Iran.

2.1. Production of kimchi

The preparation of kimchi was carried out based on the "baechu-kimchi" recipe. The ingredients are listed in Table 1. Previously, garlic, ginger, and leek were chopped. Sugar was weighed. The Chinese cabbage was cut into pieces $3 \text{ cm} \times 3 \text{ cm}$ in length and soaked in 15% (w/v) brine for 30 min.

Table 1Composition of kimchi

Materials and ingredients	Percentage share
Salted Chinese cabbage	90
Leek	4
Red Pepper powder	2
Garlic 2	2
Ginger 1	1
Sugar	1

The soaked cabbage was washed twice with fresh water and then drained for 30 min. The prepared ingredients were mixed well and then distributed evenly on the Chinese cabbage. The kimchi mixture was put into a jar. It was fermented at 20°C for 2 days and 30°C for 1 day.

2.2. Samples

In the present study, samples of kimchi were prepared. Then samples were transported to the laboratory and analyzed (Kacem etal., 2005). 25g samples were homogenized with 225mL sterile sodium citrate solution 2% (w/v), in a Stomacher 400 (Seward Medical, London, UK). Serial decimal dilutions (10^{-2} to 10^{-5}) were made in 0.1% (w/v) peptone solution.(Abdi etal.,2006) Decimal dilution of these samples were mixed with MRS medium (AEB, France) and incubated at 37°C for 48-72 h under anaerobic conditions.(Lengkey etal., 2009) The numbers of LAB were measured by the plate count on MRS agar (Difco Laboratories, Detroit, USA) mold and yeast were counted on Potato Dextrose agar (Nissui) incubated for 24 h at 30°C. Each LAB colony was purified twice by streaking on MRS agar. Colonies were counted as viable numbers of Microorganisms (colony forming unit (CFU) g⁻¹ of kimchi) (Duan etal., 2008)0.1 mL of the diluents were streaked on Nutrient agar for total bacteria counts.

2.3. Determination of pH Values and acidity of the Test Samples

The pH values of each samples were determined at 25C using a pH meter (WTW-Inolab Level 3 Terminal, Weilheim, German). (Yilmaz etal., 2010). Acid content was determined by titrating with 0.1N NaOH with the indicator of phenoluphetalin until it's colure changes to pink.

3- Results and discussion

The fermentation of kimchi is carried out through a naturally controlled brining process. During kimchi fermentation, microorganisms should be tolerant to salt, acidity, anaerobic conditions, and endogenous antimicrobial compounds in the ingredients. The main microorganisms involved in kimchi fermentation are LABs, which are facultative anaerobes, microaerobes, or anaerobes. Several factors, such as salt concentration, temperature, pH, microorganism population, and air exposure control the kimchi fermentation process. The most important factor affecting kimchi fermentation is temperature, since the kimchi fermentation occurs mainly by the microorganisms naturally present in raw materials. Kimchi is now available in all year round but the quality of kimchi differs depending on localities and seasons. Ambient temperature is applied for making kimchi at household level. Kimchi fermentation and over-acidification occurs simultaneously at ambient temperature (Yoon etal., 2000). Optimum ripening period and eatable period of Kimchi were in vestigated through panel test. Ripening time of Kimchi varied of panel test, it was evaluated that the pH of optimum ripening period of Kimchi was 4.2 respectively. The microbial changes in during laboratory fermentation of kimchi are shown in (Figure1, 2 and Table2). The LAB counts on MRS increased from 1.5×10⁴ to 1.9×10⁸ cfu/gr at 20 c. The largest increase in the numbers of LAB was noted during the first 24 h of fermentation and further incubation led to decrease. The yeast counts increased from 1.13×10^4 to 5×10^7 cfu/gr. The pH and total acid changes of kimchi are shown in (Figure 3 and 4). In the present investigation, the pH of kimchi ranged between 4.2 and 4.5. The LAB counts on MRS increased from 1.5×10^4 to 6.1×10^8 cfu/gr at 30 c. The largest increase in the numbers of LAB was noted during the first 24 h of fermentation and further incubation led to decrease. The yeast counts increased from 1.13×10^4 to 5.1×10^7 cfu/gr. The pH changes of kimchi are shown in (Figure 3 and Table 3), shows the changes of pH and total acids during kimchi fermentation at various temperatures. Ripening time of kimchi depending on fermentation temperature, accordingly the changes in pH and acidity, showed notable differences. At 20°C and 30 °C, pH dropped sharply with increasing acidity. Maximum total acid produced in kimchi at 30°C. At 30°C, the optimumripening period was 1 day and the edible period was also 1-2 days. But at lower temperature, the optimum ripening time and the edible period were longer than those of at higher temperature.

Gobbetti et al. (1994) proposed that lactic acid bacteria create an acidic environment conducive to yeast proliferation while the yeasts provide vitamins and other growth factors such as amino acids for the lactic acid bacteria. The simultaneous increase in numbers of both LAB and yeasts may therefore be attributed to their symbiotic association the results are in agreement with those reported in study. The decrease in pH and increase in lactic acid followed the same trend as reported for other natural fermented foods (Mohammed et al., 1991; Choi et al., 1994). Another reason for the difference of microflora appeared among various fermented products in

different regions was probably related to chemical and physical factors, including substrates, NaCl concentration and fermentation temperature (Cho etal., 2006; Wood, 1998).

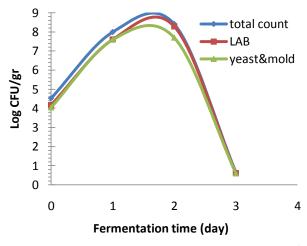


Fig. 1. Changes in M.O. during fermentation $20^{\circ c}$

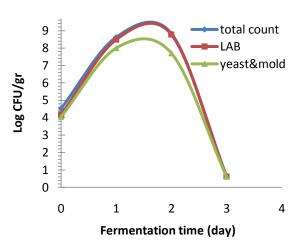


Fig. 2. Changes in M.O. during fermentation 30°c

Table 2 Change in microorganism during fermentation at $20^{\circ c}$ and $30^{\circ c}$

Yeast an	nd mold	LAB	}	Total o	ount	
30°c	20°c	30°c	20°c	30°c	20°c	day
1.13×10 ⁴	1.13×10 ⁴	1.5×10 ⁴	1.5×10 ⁴	3.4×10 ⁴	3.4×10 ⁴	0
1×10 ⁸	3.9×10^{7}	3.12×10^{8}	4×10 ⁷	4.18×10^{8}	9.9×10^{7}	1
5.1×10 ⁷	5×10 ⁷	6.11×10^{8}	1.92×10 ⁸	6.74×10^{8}	2.6×10 ⁸	2

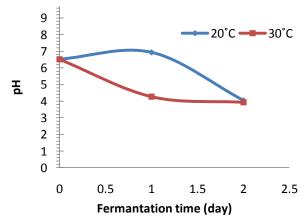


Fig. 3. Changes of total pH during Kimchi fermentation at various temperatures.

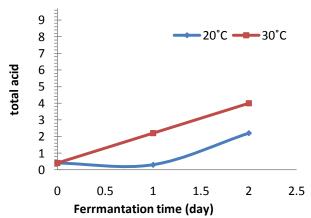


Fig. 4. Change of total acid during Kimchi fermantation at various temperatures.

Table3Change of total acid and PH during Kimchi fermentation at 20°^c and 30°^c.

Tota	Total acid		рН	
30°c	20°c	30°c	20°c	Day
0.4	0.4	6.52	6.52	0
2.2	0.3	4.27	6.93	1
4	2.2	3.93	4.05	2

References

- Arellano, R.B., Ramirez, C.I.C., Diaz, A.E., Valero, E.G., Santillan, F.M.A., 1999. Diagnosis of tuberculosis in goat flocks using the double intradermal test and bacteriology. Tecnica Pecuaria en Mexico. 37, 55-58.
- Ashraf, M., Khan, M.Z., Chishti, M.A., 1986. Incidence and pathology of lungs affected with tuberculosis and hydatidosis in sheep and goats. Pakistan Vet. J. 6, 119-120.
- Erler, W., Martin, G., Sachse, K., Naumann, L., Kahlau, D., Beer, J., Bartos, M., Nagy, G., Cvetnic, Z., Zolnir-Dovc, M., Pavlik, I., 2004. Molecular fingerprinting of *Mycobacterium bovis* subsp. *caprae* isolates from Central Europe. J. Clin. Microbiol. 42, 2234-2238.
- Francis, J., Seiler, R.J., Wilkie, I.W., O'Boyle, D., Lumsden, M.J., Frost, A.J., 1978. The sensitivity and specificity of various tuberculin tests using bovine PPD and other tuberculins. Vet. Rec. 103, 420-435.
- Grooms, D., Molesworth, J., 2000. Bovine TB Notes. Extension Bulletin E-2730 and E-2731. College of Veterinary Medicine, Large Animal Clinical Sciences, Michigan State University Extension.
- Guindi, S.M., Lofty, O., Awad, W.M., 1975. Some observations regarding the infectivity and sensitivity for tuberculosis in buffaloes in Arab Republic of Egypt. J. Egypt. Vet. Med. Assoc. 35, 125-138.
- Javed, M.T., Munir, A., Shahid, M., Severi, G., Irfan, M., Aranaz, M., Cagiola, M., 2010. Percentage of reactor animals to single comparative cervical intradermal tuberculin (SCCIT) in small ruminants in Punjab Pakistan. Acta Trop. 113, 88-91.
- Kazwala, R.R., Kambarage, D.M., Daborn, C.J., Nyange, J., Jiwa, S.F.H. Sharp, J.M., 2001. Risk factors associated with the occurrence of bovine tuberculosis in cattle in the Southern Highlands of Tanzania. Vet. Res. Commun. 25, 609-614.
- Pavlik, I., Bures, F., Janovsky, P., Pecinka, P., Bartos, M., Dvorska, L., Matlova, L., Kremer, K., Van-Soolingen, D., 2002. The last outbreak of bovine tuberculosis in cattle in the Czech Republic in 1995 was caused by *Mycobacterium bovis* subspecies *caprae*. Vet. Med. Czech, 47, 251-263.
- Pignata, W.A., Alves, C.J., Azevedo, S.S., Dantas, A.F.M., Gomes, A.A.B., Remígio, F.R., Lima, F.S., Mota, P.M.P., 2009. Prevalence for caprine tuberculosis in semi-arid of Paraíba. Pesq. Vet. Bras. 29, 7.
- Prodinger, W.M., Eigentler, A., Allerberger, F., Schonbauer, M., Glawischnig, W., 2002. Infection of red deer, cattle and humans with *Mycobacterium bovis* subsp. *caprae* in western Austria. J. Clin. Microbiol. 40, 2270-2272.
- Radostits, O.M., Gay, C.C., Blood, D.C., Hinchelift, K.W., 2000. Disease caused by bacteria *Mycobacterium*. In: *Veterinary Medicine*: A Text Book of Disease of Cattle, Sheep, Pig, Goat and Horses. 9th ed. Harcourt Publisher Ltd., London. pp. 909-918.
- Rastogi, N., Legrand, E., Sola, C., 2001. The mycobacteria: an introduction to nomenclature and pathogenesis. *Revue Scientifique et Technique de l Office International des Epizooties* 20, 21-54.
- Sanson, R.L., 1998. Tuberculosis in goats. Surveillance, 15, 7-8.
- Siva, R., William, B.J., Rao, G.D., David, W.P.A., Balasubramanian, N.N., 1997. Radiological diagnosis of respiratory diseases in large animals-a survey report. Cheiron, 26, 9-11.
- Thoen, C.O., Steele, J.H., Gilsdorf, M.J., 2006. *Mycobacterium bovis* Infection in Animals and Humans. 2nd ed. Blackwell Publishing Professional, Ames, Iowa, USA. pp. 317.
- Thorns, C.J., Morris, J.A., 1983. The immune spectrum of *Mycobacterium bovis* infections in some mammalian species: a review. Vet. Bull. 53, 543-550.
- Wood, P.R., Jones, S.L., Ellner, J.J., Brennan, P.J., Young, D., 2001. BOVIGAMTM: an in vitro cellular diagnostic test for bovine tuberculosis. Tuberculosis 81, 147-155.
- Abdi, R., Sheikh-Zeinoddin, M., Soleimanian-Zad, S. (2006). Identification of lactic acid bacteria from traditiona Iranian lighvan cheese, Pakestanjornal of biological science, 9, 99-103.

- Cho, J., Lee, D., Yang, C., Jeon, J., Kim, J., Han, H. (2006). Microbial population dynamics of kimchi, a fermented cabbage product. *FEMS microbiology letters*, 257(2), 262-267.
- Choi, S.Y., Beuchat, L.R., Perkins, L.M., Nakayama, T. (1994). Fermentation and sensory characteristics of kimchi containing potassium chloride as a partial replacement for sodium chloride. *International journal of food microbiology*, 21(4), 335-340.
- Duan, Y., Tan, Z., Wang, Y., Li, Z., Qin, G., Huo, Y., et al. (2008). Identification and characterization of lactic acid bacteria isolated from Tibetan Qula cheese. *The Journal of general and applied microbiology*, *54*(1), 51-60.
- Ekinci, R. (2005). The effect of fermentation and drying on the water-soluble vitamin content of tarhana, a traditional Turkish cereal food. *Food chemistry*, *90*(1), 127-132.
- Ekinci, R., Kadakal, C. (2005). Determination of seven water-soluble vitamins in tarhana, a traditional Turkish cereal food, by high-performance liquid chromatography. *ACTA chromatographica*, *15*, 289.
- El-Gendy, S.M. (1983). Fermented foods of Egypt and the Middle East. Journal of Food Protection 46, 358–367.
- Erbaş, M., Certel, M., Kemal Uslu, M. (2005). Microbiological and chemical properties of Tarhana during fermentation and storage as wet—sensorial properties of Tarhana soup. *LWT-Food Science and Technology*, 38(4), 409-416.
- Erdogrul, Ö., Erbilir, F. (2006). Isolation and characterization of Lactobacillus bulgaricus and Lactobacillus casei from various foods. *Turkish Journal of Biology, 30*, 39-44.
- Gobbetti, M., Corsetti, A., Rossi, J. (1994). The sourdough microflora. Interactions between lactic acid bacteria and yeasts: metabolism of carbohydrates. *Applied Microbiology and Biotechnology*, 41(4), 456-460.
- Ibanoglu, S., Ainsworth, P., Hayes, G. D. (1996). Extrusion of tarhana: effect of operating variables on starch gelatinization. *Food chemistry*, *57*(4), 541-544.
- Ibanoglu, S., Ainsworth, P., Wilson, G., Hayes, G. D. (1995). The effect o fermentation conditions on the nutrients and acceptability of tarhana. *Food chemistry*, *53*(2), 143-147.
- Lazos, E., Aggelousis, G., Bratakos, M. (1993). The fermentation of Trahanas: a milk— wheat flour combination. Plant Foods for Human Nutrition 44, 45–62.
- Lengkey, H., Balia, R., Togoe, I., Taşbac, B., Ludong, M. (2009). Isolation and identification of lactic acid bacteria from raw poultry meat. *Biotechnology in Animal Husbandry, 25*(5-6-2), 1071-1077.
- Mohammed, S. I., Steenson, L. R., Kirleis, A. W. (1991). Isolation and characterization of microorganisms associated with the traditional sorghum fermentation for production of Sudanese kisra. *Applied and environmental microbiology*, *57*(9), 2529-2533.
- Morcos, S.R., Hegazi, S.M., El-Damhougy, S.T. (1973). Fermented foods in common use in Egypt, I. The nutritive value of kishk. Journal of the Science of Food and Agriculture 24, 1153–1156.
- Salminen, S., Von Wright, A. (2011). Lactic acid bacteria: microbiological and functional aspects: CRC Press.
- Tamime, A. Y., Barclay, M. N. I., McNulty, D., O'Connor, T. P. (1999). Kishk-a dried fermented milk/cereal mixture. 3. Nutritional composition. *Le Lait*, *79*(4), 435-448.
- Wood, B. J. B. (1998). Microbiology of fermented foods (2nd ed.). London: Springer.
- Yilmaz, M.T., Sert, D., Demir, M.K. (2010). Rheological properties of Tarhana soup enriched with whey concentrate as a function of concentration and temperature. Journal of Texture Studies. 41(6):863-79.
- Yılmazer, A.N. (1994). Tarhana üretiminde kullanılabilecek uygun bir laktik asit starter kombinasyonunun geliştirilmesi (The development of favorable lactic acid starter combination for Tarhana production), Ms. Thesis, Hacettepe University, Ankara, Turkey, 83 pp.
- Sengun, I.Y., Nielsen, D. S., Karapinar, M., Jakobsen, M. (2009). Identification of lactic acid bacteria isolated from Tarhana, a traditional Turkish fermented food. International Journal of Food Microbiology. 135,105–111.