



Original article

Prevalence of helminth parasites in Gallus domesticus from Gurez valley

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ABSTRACT

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Keywords, Gurez Gallus domesticus Raillitina tetragona Ascaridia galli Heterakis gallinarum Prevalence During the study period of two years the prevalence of helminth parasites in domestic fowl from Gurez valley were analysed. A total of 137 domestic fowl were examined for helminth parasites from May 2013 to April 2015. A high rate of helminth infection (40.14%) was observed. One cestode *Raillitina tetragona* and two nematodes, *Ascaridia galli* and *Heterakis gallinarum* were encountered during the study period. High prevalence of infection was observed during summer (41.86%) followed by autumn (34.21%), spring (33.33%) and winter (30.76%). Males (36.96%) were more infected than females (34.37%). The young ones were more infected than adults. Nematodes were more prevalent than cestodes.

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1. Introduction

Gurez is a valley located in the high Himalayas on banks of river Kishenganga, about 86 km from District Bandipora and 123 km of Srinagar in northern Jammu and Kashmir, India. In northeast of Srinagar, the main valley of Gurez extends between $(34^{\circ} 30' \text{ to } 34^{\circ} 41' \text{ N} \text{ latitudes})$ and $(74^{\circ} 37' \text{ to E } 74^{\circ} 46' \text{E} \text{ longitudes})$ at an average altitude of about 2370 m.a.s.l (about 8,000 feet). It is surrounded on its north by Ladakh, by Bandipora on the south, by Ganderbal on its southeast and on the west by Kupwara with its peripheries touching Line of Control (LoC) that divides the states of India and Pakistan. The valley is nestled among high towering peaks and lofty and glaciated snow capped mountains which are not just an unvarying landmass but show great differences in elevation aspect, rock type, ruggedness and glacial work which coalesce to make contrasting land surfaces. One

has to cross the coldest and dangerous peak Razdan (Razdan pass) located above 4000 m.a.s.l. to reach the valley. The pass not only connects the region with the rest of Kashmir but also divides the two on geographical, sociocultural and linguistic lines. The valley is drained by mighty Kishenganga (Neelam) River between Kaobal Gali in east and Kanzalwan in west while other aquamarine and crystal streams also run through it.

Birds are important for their commercial, recreational, ethical, spiritual values and form a rich protein source for humans. There is a rich diversity of birds in this area. Among birds, Poultry is of great importance in rural production system in this area. Chicken was the only poultry bird being reared for meat and egg production. Unlike rest of Bandipora district, which has a good population of duck and geese, no non-chicken poultry species was reported from the area. The average number of chicken per household varied from 5- 10. During summer months, hens laid as many as 20 eggs/month whereas no egg production was reported during winter months; which may be attributed to the short day length coupled with very little scavenging feed resources. The domestic fowl and eggs provide an important source of protein for human consumption. The increased mortality and decreased productivity in chickens is mainly due to mismanagement, lack of nutritional feeding, diseases and predation.

Among the problems facing extensive types of production of chickens in Gurez are parasitic diseases. There are numerous species of helminthes that cause significant damage to the organs in which they live. Cestodes are more commonly found in warm weather when intermediate hosts are abundant. Helminthiasis was considered to be important problems in chickens (Jansen and Pandey 1989) and (Abebe *et al.*, 1997). Helminth parasites were increminated as major causes of unsoundness and lowering performance of poultry in Egypt (Khater, 1993). Avian helminthiasis constitutes one of the most common endoparasitism causing serious troubles in chicken production. Chicken cestodiasis not only cause loss of body weight of the raised chickens but also may cause several problems in affected flocks such as enteritis, loss of blood, loss of production, nervous manifestations and death (Calneck *et al.*, 1997). The prevalence and intensity of helminth infections in birds may be influenced by several factors as distribution of intermediate hosts such as beetles, ants, crustaceans, houseflies *etc.* and their infection rate and the number of infective parasite eggs or larvae. The free ranging management system and climatic conditions, such as temperature and humidity alter the population dynamics of parasites resulting in dramatic change in prevalence and intensity of helminth infections.

2. Materials and methods

During the present study from May 2013 to April 2015, the domestic fowl were purchased from the local people at different study sites. The hosts were then taken alive to the temporary laboratory maintained at Dawarthe capital of Gurez or were brought alive to Department of Zoology, University of Kashmir for parasitic examination. For the collection of endoparsites the body of the hosts was dissected open midventrally and different organs including alimentary canal were removed and kept in separate desired size petridishes where these organs were teased and cut open to search for parasites if any. Therefore, the gastrointestinal tract was subjected to routine examination to collect the gastrointestinal parasites, according to the procedure as described by Fowler (1990). Cestodes were collected by the help of dropper and preserved in 10% formalin or cornoy's fluid for the identification. Morphology of cestodes was studied by preparing permanent slide according the methods as described by Cable (1957). After washing, nematodes were collected by the help of curved needle and kept in glycerin alcohol. Nematodes were best killed in steaming hot 70% alcohol, and stored in the same solution. Later, a few drops of glycerin were added. Thorough morphological study of nematodes was performed by the preparation of sub-permanent slide by adding one drop of lactophenol. The other steps in this were fixation, staining, dehydration, de-alcoholisation and clearing, mounting and labeling. Parasites were identified according to the keys and description given by Soulsby (1982) and Yamaguti (1958). On comparing the recovered parasites were identified as Raillitina tetragona, Ascaridia galli and Heterakis gallinarum.

Light microscopy was conducted under Olympus Research microscope with lens combination of 7X, 10X, and 15X eye pieces and 4X, 10X, 20X, 40X and 100X objectives. The drawings for identification purposes were made to scale with the help of prism type camera-Lucida. The Photographs were taken with the help of Sony Digital SLR Camera Model Number (DSLR – A200). Photomicrography was conducted with DP – 12 Digital Camera attached to Olympus Research Microscope in the department of Zoology.

The prevalence of Helminthiasis was recorded as per formulae described by Thrusfield (1986).

Prevalence =	No. of individual having a disease at a particular point in time ×100				
	No. of individual in the population at risk at that point in time				
	Definitions				
	The ecological terms used in this study are				
Prev	alence = Total number of hosts infected x 100				
	Total number of hosts examined				
Mean Intensity =	Total number of Parasites				
-	Total number of hosts infection				
	Total number of nosts infection				
Relative Density or Abundance =	Total number of Parasites				
	Total number of hosts examined				

The above nomenclature is followed by that given by Morgolis et al. (1982).

2.1. Data analysis

The most common measurements of parasite population levels in hosts are prevalence, mean intensity and mean abundance Bush et al. (1997). Prevalence refers to the percentages of organisms infected by a particular species of parasite. Mean intensity is the number of parasites of a given species per infected host. Mean abundance refers to the number of parasites of a given species per host examined, infected and uninfected. The nomenclature used to define ecological parameters is in consistency with that of Margolis et al. (1982).

3. Results and discussion

The three different helminth parasites recovered belonging to two classes; cestoda and nematode were observed during the present study.

3.1. Prevalence

A total of 137 specimens of fowl were examined during the present study which revealed 40.14% (55/137) of infection by helminthes in this beautiful valley. Three different types of helminth parasites recovered during the study include two nematodes (*Heterakis gallinarum & Ascaridia galli*) and one cestode i.e., *Raillietina tetragona*. *Heterakis gallinarum* showed a highest prevalence of 35.76% followed by *Ascaridia galli* (32.11%) and *Raillietina tetragona* (27.00%) (Table 1 & 2).

Host	NE	Un- infected	Infected	%age	Trematode	Cestode	Nematode	
Fowl	137	82	55	40.14%		37 (27.00%)	49 (35.76%)	
Table 2 Prevalenc	e of helminth	es.						
Host infe	cted with	No.	examined	No.	infected	%a	ge	
Raillietina	tetragona		137		37		27.00%	
Ascaridia	galli				44	32.1	1%	
Heterakis	gallinarum				49	35.7	6%	
			T-Valu	e = 26.91 P-	Value = 0.001			

Table 1

3.1.1. Seasonal prevalence

The study showed that the prevalence of parasites in fowl was throughout the year but the prevalence varied from season to season. The highest prevalence was observed during summer followed by autumn and least in winter. During summer 43 fowl were examined, out of which 15 (34.88%), 17 (39.53%) and 18 (41.86%) were found infected with *Raillietina tetragona, Ascaridia galli* and *Heterakis gallinarum* respectively. Similarly during autumn out of 38 specimens examined, 9 (23.68%), 12 (31.57%) and 13 (34.21%) were infected with *Raillietina tetragona, Ascaridia galli* and *Heterakis gallinarum* respectively. However a lowest prevalence of these helminth parasites was observed during winter. Out of 26 specimens examined 5 (19.23%); 6 (23.07%) and 8 (30.76%) were infected with *Raillietina tetragona, Ascaridia galli* and *Heterakis gallinarum* respectively (Table 3). Thus a decreasing order of prevalence was summer>autumn>spring>winter.

Host	Season	NE	Infected		
			Raillietina tetragona	Ascaridia galli	Heterakis gallinarum
	Spring	30	8 (26.66%)	9 (30.00%)	10 (33.33%)
Fowl	Summer	43	15 (34.88%)	17 (39.53%)	18 (41.86%)
	Autumn	38	9 (23.68%)	12 (31.57%)	13 (34.21%)
	Winter	26	5 (19.23%)	6 (23.07%)	8 (30.76%)
Total		137	37 (26.11%)	44 (31.08%)	49 (35.04%)
			T-Value = 12.25 P-	T-Value = 14.52	T-Value = 12.36 P-
			Value = 0.001	P-Value = 0.001	Value = 0.001

Table 3

3.1.2. Age-wise prevalence

Fowl specimens of different age groups were examined. Out of 49 examined fowl specimens of 0-6 months age, 14 (28.57%), 17 (34.69%) and 18 (36.73%) were infected with *Raillietina tetragona, Ascaridia galli* and *Heterakis gallinarum* respectively. Similarly out of 54 fowl specimens of 6 months - 2 years age group, 15 (27.77%), 18 (33.33%) and 19 (35.18%) were infected with *Raillietina tetragona, Ascaridia galli* and *Heterakis gallinarum* respectively. However out of 34 hosts from 2 years and above age group only 8 (23.52%), 9 (26.47%) and 12 (35.29%) were infected with *Raillietina tetragona, Ascaridia galli* and *Heterakis gallinarum* respectively. The results indicate that the hosts have maintained a moderate resistance against cestodes with advancement of age. However there is no significant age resistance shown by the hosts against nematode infection. Thus the hosts of any age group may be exposed to helminth infections with a slight resistance developing during the advancement of age.

				Cestode	Nema	tode
Host	Age group	NE	Trematode	Raillietina tetragona	Ascaridia galli	Heterakis gallinarum
	<6 month	49		14 (28.57%)	17 (34.69%)	18 (36.73%)
Fowl	6 months-2 years	54		15 (27.77%)	18 (33.33%)	19 (35.18%)
	>2 years	34		8 (23.52%)	9 (26.47%)	12 (35.29%)
Total		137		37 (26.62%)	44 (31.49%)	49 (35.73%)
				T-Value = 8.67	T-Value = 9.64	T-Value = 7.63
				P-Value = 0.013	P-Value = 0.011	P-Value = 0.017

3.1.3. Sex-wise prevalence

Out of 137 specimens of *Gallus domesticus* examined during the present study, 73 were males and 64 were females. A prevalence of 28.76% (21/73), 32.87% (24/73) and 36.96% (27/73) in males, and 25% (16/64), 31.25% (20/64) and 34.37% (22/64) in females of *Raillietina tetragona, Ascaridia galli* and *Heterakis gallinarum* respectively was observed during the present study (Table 5 & 6). The results show that there is no marked but a slight resistance shown by females as compared to males.

Table 5

Sex-wise prevalence of helminthes.

Host Sex		NE	Trematode	Cestode	Nema	Nematode	
				Raillietina	A. galli	H. gallinarum	
				tetragona	0.	0	
Fowl	Male	73		21 (28.76%)	24 (32.87%)	27 (36.96%)	
	Female	64		16 (25.00%)	20 (31.25%)	22 (34.37%)	
Total		137		37 (27.16%)	44 (32.25%)	49 (35.76%)	
				T-Value = 47.40	T-Value = 57.50	T-Value = 45.00	
				P-Value = 0.013	P-Value = 0.011	P-Value = 0.014	

Table 6

Single and multiple species infection in domestic fowl.

	NE	Parasite	NI	%age
	137	Raillietina tetragona	21	15.32
		Ascaridia galli	19	13.86
		Heterakis gallinarum	26	18.97
Multiple type		Raillitina + Heterakis	5	3.64
		Raillitina + Ascaridia	7	5.10
		Ascaridia + Heterakis	14	10.21
		Raillitina+Heterakis+Ascaridia	4	2.91

After extensive study of the bird host, Gallus domesticus for helminth parasitism in Gurez valley during the present study, different species of helminth parasites were recovered with a moderately high prevalence (40.14%). These results when compared with those of other researchers around the globe indicate that this small, however isolated, valley does not figure out of the helminth infestation; but is an endemic area for helminth parasites in fowl. Permin and Nansen (1996) after studying Danish organic poultry farming reported increased infestation of internal parasites including Heterakis gallinarum. The author further stated that species of nematodes like Heterakis and Ascaridia are widely distributed, causing non-specific clinical signs of infection, such as loss of appetite and growth, a general in condition and on occasions death. Permin et al (1999) in their survey in Denmark concluded that there was a high risk of helminth infection in free range/ organic poultry systems and that prevalence may also be high in deep litter systems. Permin et al (1997) reported that prevalence of GIH are high whether in tropical or temperate climate. Oyeka (1989) found 54.5% of chickens to be infected with helminth parasites in Anambra state in Nigeria. Yadav and Tandon (1989) revealed 90.9% of helminth infections in subtropical high rainfall area of India. Mpoame and Agbede (1995) found 93.55% of domestic fowl infected with gastrointestinal helminthes Eshetu et al (2001) found 91.01% chickens infected with gastrointestinal helminthes from Amhara region Ethiopia.Nokana et al (1991) during their survey of helminth parasites inbackyard flocks in Michigan by litter examination also showed relatively high contamination rates. Edger (1953) reported the presence of a wide range of helminthes in chickens including A. galli. Wilson et al (1994) revealed the prevalence of A. galli was in the range of 40% on commercial farms in the state of Arkansas. Long (1977) reported that majority of broiler chickens were infected with A. galli at the age of 23-29 days. Konanenko and Khaizade (1983) while working on helminth fauna of Charadriiformes and Anseriformes observed a prevalence of 50%, 45%, 13.3% and 16% of cestodes, nematodes, trematodes and acanthocephalans respectively. Luka and Ndams (2007) reported that 62% of domestic fowl were infected with helminth parasits in Samaru, Zaria Nigeria. Phiri et al (2007)

reported 95.2% free range chickens infected with GIH in central Zambia.Wahid *et al* (2006) reported a prevalence of 30.75%, 31.25% of *Raillitina* spp. and *Heterakis* spp. in domestic ducks, and a prevalence of 50% of *Ascaridia galli* in goose from Srinagar Kashmir. Ayesha *et al* (2007) reported a prevalence of 51.42%, 31.42% and 30.71% of *Raillitina tetragona, Heterakis gallinarum* and *Ascaridia galli* in domestic fowl from Doda district of Jammu and Kashmir.

The remarkable prevalence of infection observed in domestic fowl from Gurez valley can be attributed to a number of factors like the type of management and production system, exposure to intermediate hosts, inadequate or no use of anthelmintics, the climatic conditions which alter the population dynamics of the parasite. As the environment and climatic conditions of the study area (Gurez valley) seem to be unfavourable for such a high intensity of infection that could have as such resulted in a lowest level of infection, a major ecological factor like dispersal and emigration plays a significant role in maintaining such a high prevalence of helminth infection in this beautiful valley of Gurez. During severe winters a small fraction of human population along with their poultry birds migrate from this valley to Kashmir valley until the onset of spring. Thus a chance of transmission of infection (even if minimum) might have occurred which shows a good proliferation while the birds are back in their native land. During pleasant summers, the shepherds, Bakerwals along with their poultry birds, visit the high altitude pastures of Gurez for grazing their sheep and goats where the possible chances of transmission of helminth infection was observed during summers.

In the present study, a good number of domestic fowl were harbouring more than one type of helminth species which is in agreement with the work of many other researchers like Edger (1953), Konanenko and Khaizade (1983), Qureshi (1950), Islam *et al* (1988), Yadav and Tandon (1989), Wilson *et al* (1994), Mpoame and Agbede (1995), Permin and Nansen (1996), Permin *et al* (1999), Eshetu *et al* (2001), Magwisha *et al* (2002), Fakae and paul-Abiade (2003), Hossouni and Beighti (2006), Schou *et al* (2006), Phiri *et al* (2007), and Luka and Ndams (2007).

The most prevalent helminth parasite recorded during the present study from Gurez was *Heterakis* spp. (35.76%) followed by *Ascaridia galli* (32.11%) and *Raillitina* spp. (27.00%). The present study is in agreement with the work of many others like; Qureshi (1950) reported a high prevalence of *Ascaridia galli* (31.02%) and *Raillitina tetragona* (18.7%) in Desi adult fowls in U.P. India. Wilson *et al* (1994), Permin *et al* (1999) in deep litter and backyard system, Eshetu *et al* (2001) from Amhara region also reported similar prevalence in their studies. However Mpoame and Agbede (1995), Schou *et al* (2006), Luka and Ndams (2007) reported a higher prevalence of *Ascaridia galli*. Since the present results are in agreement with those of many others, still the variations can be attributed to the environmental conditions in the area and inadequate availability of intermediate hosts. The environmental conditions like temperature and moisture do favour the larval development and facilitate transmission and ingestion of infested droppings.

The present studies indicate that with the advancement of age there was a decrease in the prevalence of infection which makes it in agreement to the reports made by Soulsby (1982), Negesse (1991), Sanders and Schwartz (1994), Mpoame and Agbede (1995), Magwisha *et al* (2002), and Phiri *et al* (2007). It is evident that it can be attributed to the increased immune status in adults than in young ones against the helminth parasites. To accommodate with the environmental conditions and thus traditionally the chicks do hatch out during late spring and as such they do enjoy the summer and autumn in their young age when these get exposed to helminth infections. These could be the possible reasons for decrease in prevalence of helminth infection with increase in age.

As is evident from the observations made during the present study that males do show a slightly higher rate of infection than females, which can be related with the physiological influence of hormones on the susceptibility of host animals to infection (Magwish *et al* 2002). Similar trend was observed by Fakae and paul-Abiade (2003) in fowl where male fowls carried significantly (p<0.05) more parasite burden than females. Magwish *et al* (2002) also observed that prevalence of *Heterakis gallinarum* was higher in males than females. Similar results were observed by Ayesha *et al* (2007) in domestic fowl from Doda district of Jammu and Kashmir.

Although helminth parasites were prevalent throughout the year, but the observations made in the present study show higher prevalence of infection during the warm summer months followed by autumn, spring and winter respectively. As the study area is far-off from University of Kashmir and along the Line of control, monthly investigations were not possible. Thus, seasonal investigations were made to study the helminth infestation throughout the year. The higher prevalence observed during summer and lower during cold winters can be due to the impact of many factors like geographical location of the area, environmental conditions prevailing in the area.

Low temperature inhibits the development and survival of infective larval stages and as such decreases the access to intermediate hosts or final hosts. On the other hand, the enough availability of intermediate hosts and favourable temperature for larval development favours the chances of helminth proliferation in summers. It can be suggested that the seasonal fluctuations in the abundance of infective stages in the environment may also play a contributing role in the differences observed. These findings are in accordance to the reports made by many workers from other parts of the world. Threlfall (1967) examined herring gulls in north-wales and found seasonal diversity of helminth parasites and attributed the differences to changes in the diet of the gulls over the course of the year caused by an altered availability of the intermediate hosts. Busher (1965) and Mclaughlin and Burt (1979), reported that the density and the magnitude of helminth infection increases to peak in late summer. Wallace and Pence (1986) stated that there is no recruitment of replacement species during winters due to changes in diet and limited availability of infective stages in intermediate hosts. Fedynich and Pence (1994) reported that mallards had higher mean abundance of helminth in summer than in winter. Mpoame and Agbede (1995) reported that the parasitic prevalence and the worm burdens were generally higher during April to October. Magwisha *et al* (2002) observed that helminth infection varied in the months of rainy season.

The present study reveals that single type infections were more prevalent than multiple type infections. Multiple type infections with helminthes in domestic fowl was also observed by Yadav and Tandon (1991), Mpoame and Agbede (1995), Permin et al (1997), Magwisha et al (2002), and Phiri et al (2007). In this study, majority of the host birds harboured multiple type of infection of helminthes which suggest that the prevailing environmental conditions and free range management systems are favourable to many species of helminth parasites.

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